

Regenerative Medicine Combined with Physiotherapy in Orthopedic Rehabilitation: Evidence, Mechanisms, and Clinical Protocols



*Correspondence:

P. Muthukrishnan, M.P.T (Ortho), Ph.D. Scholar, Meenakshi Academy of Higher Education & Research (MAHER), University in Chennai, Tamil Nadu,

E-mail: krishphysio5335@gmail.com
Received Date: 09 Dec 2025
Accepted Date: 16 Dec 2025
Published Date: 18 Dec 2025

Citation:

Muthukrishnan P, Rajadurai S.
Regenerative Medicine Combined
with Physiotherapy in Orthopedic
Rehabilitation: Evidence, Mechanisms,
and Clinical Protocols. WebLog J
Orthop. wjor.2025.11801. https://doi.
org/10.5281/zenodo.18055250

Copyright© 2025 P. Muthukrishnan.

This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

P. Muthukrishnan1* and S. Rajadurai2

¹Meenakshi Academy of Higher Education & Research (MAHER), University in Chennai, Tamil Nadu, India

²Associate Professor, Department of Orthopedics, Meenakshi Medical College Hospital and Research Institute, Kanchipuram, Tamil Nadu, India

Abstract

Regenerative medicine encompasses emerging biologic interventions including platelet-rich plasma (PRP), mesenchymal stem cell (MSC) therapies, and tissue engineering approaches that harness growth factors and cellular biology for musculoskeletal repair. While individually effective, recent evidence demonstrates that combining regenerative therapies with evidence-based physiotherapy protocols produces superior functional outcomes, sustained pain relief, and structural tissue regeneration compared to monotherapy approaches. This evidence-based review examines the synergistic mechanisms, clinical efficacy data, and integrated rehabilitation frameworks that optimize outcomes in orthopedic tissue disorders. A comprehensive scoping review of recent clinical trials and meta-analyses reveals that combined regenerative medicine with progressive exercise therapy significantly improves pain, function, and quality of life in knee osteoarthritis, rotator cuff pathology, tendon injuries, and cartilage defects. Critical gaps persist regarding standardized treatment protocols, long-term cost-effectiveness, optimal timing of physiotherapy initiation, and biomarkers predicting individual responder status. This review provides clinically relevant evidence supporting the integration of physiotherapy with biologic interventions, emphasizes mechanisms of synergy between growth factor delivery and mechanical loading, and highlights future research priorities essential for advancing precision medicine approaches in orthopedic rehabilitation.

Keywords: Regenerative Medicine; Platelet-Rich Plasma; Mesenchymal Stem Cells; Physiotherapy; Tissue Engineering; Growth Factors; Orthopedic Rehabilitation; Non-Surgical Management

Introduction

The orthopedic field faces substantial clinical challenges in managing musculoskeletal injuries and degenerative conditions. Traditional approaches have historically emphasized symptom management through conservative care or surgical intervention, yet both modalities possess inherent limitations. Conservative therapy, while cost-effective, often provides incomplete healing and high recurrence rates in complex injuries. Conversely, surgical intervention carries risks of infection, extended rehabilitation periods, and diminished long-term functional outcomes when applied to certain conditions.

Recent advances in regenerative medicine have fundamentally shifted the therapeutic paradigm. Biologic interventions including platelet-rich plasma (PRP), mesenchymal stem cell (MSC) therapies, bone marrow aspirate concentrates, and tissue engineering scaffolds harness the body's intrinsic healing mechanisms to promote durable tissue regeneration rather than merely managing symptoms. These therapies deliver high concentrations of bioactive growth factors including transforming growth factor-beta (TGF- β), bone morphogenetic proteins (BMPs), insulin-like growth factor-1 (IGF-1), vascular endothelial growth factor (VEGF), and platelet-derived growth factor (PDGF) directly to sites of tissue damage [1, 2].

However, mounting clinical evidence reveals a critical gap in current practice: biologic therapies achieve substantially greater efficacy when integrated with structured physiotherapy than when administered as monotherapy. Approximately 30-40 percent of patients demonstrate inadequate response to regenerative interventions alone, suggesting the necessity of adjunctive rehabilitation

strategies to optimize tissue remodeling and functional recovery [3]. The synergistic interaction between biologic therapy and mechanical loading through exercise represents an underexplored yet increasingly validated mechanism for maximizing orthopedic outcomes.

Contemporary orthopedic practice increasingly emphasizes nonsurgical, minimally invasive approaches. This shift reflects growing evidence supporting early combined intervention, expanding patient demand for non-operative options, and accumulating data demonstrating cost-effectiveness of non-surgical pathways when properly integrated [4, 5]. Physiotherapy combined with biologic therapy bridges the gap between conservative management and invasive surgery, providing a sophisticated middle approach that addresses underlying pathophysiology while avoiding surgical morbidity.

This evidence-based review synthesizes current literature on regenerative medicine combined with physiotherapy, examines mechanisms of synergy between growth factors and mechanical loading, evaluates clinical efficacy data across musculoskeletal conditions, proposes integrated rehabilitation protocols, and identifies critical research gaps requiring investigation.

Mechanisms of Synergy: Growth Factors and Mechanical Loading

Growth Factor Biology and Tissue Repair

Biologic therapies exert therapeutic effects through delivery of growth factors that initiate and sustain tissue regeneration cascades. Transforming growth factor-beta (TGF- β) superfamily members constitute the most extensively investigated growth factors in cartilage and bone repair. TGF- β 1 stimulates chondrocyte synthetic activity while decreasing catabolic activity mediated by inflammatory cytokines including interleukin-1 (IL-1) [6]. *In vitro* and animal studies demonstrate that TGF- β 1 and TGF- β 3 enhance chondrogenic differentiation of bone marrow-derived mesenchymal stem cells and promote extracellular matrix synthesis with composition resembling hyaline cartilage rather than fibrocartilage [7, 8].

Bone morphogenetic proteins, particularly BMP-2 and BMP-7 (osteogenic protein-1), possess potent osteogenic and chondrogenic effects. BMP-7 currently represents the gold standard growth factor for articular cartilage repair based on cumulative evidence [9]. These proteins stimulate matrix synthesis while simultaneously suppressing catabolic mechanisms including matrix metalloproteinase expression and inflammatory cytokine production. Synergistic effects emerge when multiple growth factors are combined; studies demonstrate that BMP-7 combined with insulin-like growth factor-1 produces greater cartilage matrix synthesis than either factor alone [10].

Fibroblast growth factors (FGF), particularly FGF-2 and FGF-18, contribute to bone and cartilage regeneration by promoting osteoblast function, soft callus formation, and mesenchymal stem cell proliferation [11]. Vascular endothelial growth factor (VEGF) drives angiogenesis essential for supplying oxygen and nutrients to regenerating tissue while recruiting circulating progenitor cells [12]. Platelet-derived growth factor (PDGF), abundant in platelet-rich plasma preparations, enhances cell migration, proliferation, and recruitment of reparative cells [13].

The efficacy of growth factor therapies depends critically upon delivery methodology. Bioactive scaffolds incorporating controlledrelease systems provide sustained growth factor availability over weeks to months, achieving spatial and temporal patterns mimicking physiologic tissue development [14]. Three-dimensional tissue engineering platforms utilizing polyglycolic acid (PGA), polylactic acid (PLA), and biodegradable composite scaffolds create microenvironments supporting cell survival, differentiation, and extracellular matrix deposition while guiding tissue morphogenesis toward anatomically functional architectures [15].

Mechanical Loading and Cellular Response

Mechanical loading through exercise represents a critical but often underappreciated stimulus for tissue regeneration. Mechanical forces trigger mechanotransduction—cellular processes converting physical stimuli into biochemical signals—through multiple mechanisms [16]. Chondrocytes and tenocytes respond to mechanical loading by upregulating synthesis of growth factors including TGF- β and IGF-1, essentially creating an endogenous growth factor milieu amplifying exogenously delivered biologic agents [17].

Progressive resistance exercise stimulates mesenchymal stem cell proliferation and differentiation through multiple pathways. Mechanical tension activates intracellular signaling cascades (involving MAPK, PI3K/Akt pathways) promoting osteogenic and chondrogenic differentiation depending upon loading parameters [18]. Appropriate mechanical stimulation also enhances collagen synthesis and promotes longitudinal alignment of collagen fibers, critical for developing tensile strength and functional load-bearing capacity in repaired tissues [19].

Aerobic exercise and non-impact cardiovascular training enhance angiogenesis through both mechanical and metabolic mechanisms, increasing blood flow and delivering nutrients and circulating progenitor cells to regenerating tissues [20]. Neuromotor exercises restoring proprioception and motor control optimize biomechanical loading patterns, reducing aberrant stress concentrations and redistributing loads across regenerated tissues more evenly [21].

Synergistic Mechanisms

The synergy between biologic therapy and physiotherapy emerges from complementary mechanisms. Growth factors create a biological permissive environment favoring tissue regeneration, yet without mechanical guidance, this regeneration often produces mechanically inferior fibrocartilage or disorganized tissue architecture [22]. Conversely, mechanical loading alone in damaged tissues with compromised healing capacity produces inadequate regenerative response.

The integrated approach leverages growth factors to accelerate cellular proliferation and differentiation while mechanical loading through progressive physiotherapy:

- Optimizes collagen fiber alignment and crosslinking.
- Enhances vascular integration through mechanically-stimulated angiogenesis.
- Activates endogenous growth factor production through mechanotransduction.
 - Restores functional motor patterns and proprioceptive control.
 - Prevents maladaptive fibrosis and contracture formation.

Recent scoping reviews examining combined regenerative medicine with exercise therapy in knee osteoarthritis demonstrate significantly greater improvements in pain and function than regenerative medicine monotherapy [23, 24]. Patients receiving PRP injections alongside supervised progressive exercise showed sustained pain relief and functional improvements extending beyond 96 weeks, suggesting durable tissue regeneration. The timing of physiotherapy initiation relative to biologic injection represents a critical optimization variable; evidence supports initiating gentle active motion by weeks 2-3 post-injection and progressing resistive loading by weeks 6-8 pending individual healing response and imaging confirmation of tissue integrity [25].

Clinical Efficacy in Musculoskeletal Disorders

Knee Osteoarthritis

Knee osteoarthritis represents the most extensively investigated condition for combined biologic and physiotherapy interventions. Meta-analytic evidence involving 1,995 patients across randomized controlled trials demonstrates that platelet-rich plasma provided statistically superior improvements in VAS pain scores (mean difference -1.8; 95% CI: -2.1 to -1.5) and WOMAC functional scores compared with placebo at all follow-up timepoints [26, 27]. Critically, improvements exceeded the minimal clinically important difference (MCID) at 3- and 6-month follow-up points for VAS pain, and at all follow-up points for WOMAC functional scores [28].

A recent scoping review identified three randomized trials directly examining combined regenerative medicine with exercise therapy in knee osteoarthritis. All three studies demonstrated significantly greater improvements in pain and function in combined intervention groups compared to regenerative medicine monotherapy [29]. These improvements persisted both short-term (6-24 weeks) and long-term (up to 96 weeks), suggesting additive or synergistic treatment effects. The mechanisms underlying superior outcomes involved complementary physiologic processes: regenerative therapy provided biological substrate for tissue repair while progressive exercise optimized mechanical properties and functional integration of regenerated cartilage [30].

Subgroup analyses comparing mesenchymal stem cell therapy with PRP revealed stem cell therapy produced greater efficacy in pain reduction and functional improvement, attributed to superior immunomodulatory properties and enhanced capacity for autologous tissue regeneration [31]. However, variability in MSC sourcing (bone marrow-derived versus adipose-derived), cell dosage, and preparation protocols significantly influenced outcomes. Studies employing higher MSC dosages (≥50 million cells) and repeated intra-articular injections achieved superior functional improvements documented by MRI-T2 mapping and radiographic assessments showing improved joint space width and subchondral bone structure [32, 33].

Rotator Cuff Pathology

Rotator cuff tendinopathy and partial-thickness tears represent prevalent conditions in both athletic and sedentary populations. The inherent challenge in rotator cuff healing stems from limited vascularization of tendinous tissue, restricted capacity for self-repair, and substantial mechanical demands during shoulder function [34]. Systematic reviews examining platelet-rich plasma for rotator cuff tendinopathy demonstrate mixed results regarding long-term efficacy. Short-term benefits emerge prominently: meta-analytic evidence shows PRP provides statistically significant improvements in pain reduction and functional scores through 6-month follow-up

compared to placebo and corticosteroid injections [35]. However, PRP's efficacy tends to diminish beyond 6 months, often becoming comparable to placebo by 12-month follow-up in isolated biologic therapy studies [36].

Integration of structured physiotherapy fundamentally alters this trajectory. Comprehensive rehabilitation protocols for rotator cuff pathology emphasize sequential progression from scapular stabilization exercises and rotator cuff muscle activation in early phases, progressing to resistance training and dynamic stability exercises, culminating in sport-specific or occupational activity simulation [37, 38]. Exercise programs targeting high middle and lower trapezius activation with minimal upper trapezius participation have demonstrated superior neuromuscular outcomes. Studies incorporating combined PRP injection with progressive physiotherapy show sustained functional improvements through 12-month follow-up and reduced surgical conversion rates (approximately 9-15% at 12 weeks compared to historical surgical conversion rates exceeding 30-40% in conservative management alone) [39].

Tendon and Ligament Injuries

Biologic therapies show particular promise in tendon pathology given the limited healing capacity of avascular tendinous tissue. Stem cell-based strategies, particularly bone marrow-derived mesenchymal stem cells (BMSCs) and tendon-derived stem cells (TDSCs), demonstrate capacity for enhanced tendon regeneration when combined with appropriate scaffolding and growth factor delivery [40, 41]. TDSC-based approaches exploit the cells' inherent ability to differentiate into tenocytes, osteoblasts, and chondrocytes, producing both the tendon substance and the enthesis (tendon-bone interface) critical for successful healing [42].

Growth factor combination therapies show enhanced efficacy: delivery systems combining transforming growth factor-beta and ghrelin produced superior cartilage and tendon regeneration compared to single growth factors [43]. Dual-delivery systems using microsphere/hydrogel matrices enabling sustained release of TGF- β 3 and ghrelin significantly enhanced chondrogenic differentiation and tissue formation [44].

Physiotherapy integration proves essential in ligament injuries. While PRP demonstrates potential for accelerating graft maturation (assessed by MRI signal intensity changes) and reducing postoperative effusion, most studies show PRP monotherapy insufficient for improving long-term functional outcomes, ligament laxity, or graft failure rates [45, 46]. Combined approaches incorporating anterior cruciate ligament reconstruction with periligamentous PRP injection plus structured progressive rehabilitation demonstrate superior return-to-sport outcomes [47]. Current evidence supports initial relative rest (1-2 days post-injection) followed by progressive activity initiation. By weeks 2-3, controlled active motion commences; weeks 4-5 introduce exercise progressions pending imaging confirmation of tissue integrity; weeks 6-8 initiate eccentric loading; weeks 8-10 progress to plyometrics and sport-specific activities; return to sport occurs at weeks 12-15 [48].

Integrated Rehabilitation Protocols

Effective integration of physiotherapy with regenerative medicine requires temporal coordination, appropriate exercise progression, and physiologic understanding of tissue healing timelines. The following protocol framework, informed by current evidence and clinical experience, provides a foundation for tissue-specific

Table 1:

Phase	Timeline	Primary Physiotherapy Focus	Exercise Progression	Integration with Biologic
Phase 1	Weeks 0-2	Protection, ROM restoration	Passive/active-assisted motion	Growth factor initiation; inflammation management
Phase 2	Weeks 3-6	Active motion, tissue mobilization	Light resistance, neuromuscular control	Cellular differentiation; angiogenesis; matrix synthesis
Phase 3	Weeks 7-12	Functional loading, sport-specific	Resistance training, plyometrics	Collagen organization; mechanical adaptation
Phase 4	Weeks 13+	Return to sport/activity	Unrestricted functional activities	Tissue maturation; long-term durability

applications (Table 1).

Phase 1 (Weeks 0-2): Protection and Foundational Motion

Immediate post-injection management prioritizes injection site protection and inflammation management. Patients maintain relative rest of the affected region for 1-2 days, allowing medication absorption and monitoring for adverse reactions [49]. Ice application (15 minutes, 3-4 times daily) during the first week reduces reactive inflammation and swelling.

Physiotherapy interventions emphasize restoration of passive and active-assisted range of motion within pain-free parameters. Manual therapy techniques address myofascial restrictions developing from protective muscle guarding. Modalities including cryotherapy, transcutaneous electrical nerve stimulation (TENS), and interferential current manage pain and facilitate inflammation resolution. By week 2, achievement of 90 percent full range of motion represents a key progression criterion [50].

Biologic considerations: Growth factors released from platelets (in PRP) or from infused stem cells initiate chemokine signaling recruiting endogenous reparative cells. Gentle motion prevents tissue adhesion formation while promoting early neovascularization [51].

Phase 2 (Weeks 3-6): Active Motion and Tissue Remodeling

Progression criteria from Phase 1 include minimal swelling, absence of joint effusion on clinical examination, full passive range of motion, and pain controlled to VAS 4-5 or less. Phase 2 emphasizes active movement with progressive loading integration.

Physiotherapy interventions progress to active-range-of-motion exercises, initially against gravity alone, advancing to light resistance by week 4-5 [52]. Soft tissue mobilization techniques, including instrument-assisted soft tissue mobilization and gentle myofascial release, enhance tissue extensibility and blood flow. Proprioceptive training and early neuromuscular control exercises commence. Cardiovascular conditioning utilizing non-weight-bearing modalities (swimming, aquatic therapy, upper body ergometry) maintains aerobic capacity without imposing excessive joint loads.

By week 6-7, patients should achieve substantially improved pain (VAS 2-3), full range of motion, and capability for moderate resistance training pending imaging confirmation of tissue integrity via ultrasound or MRI [53].

Biologic considerations: Growth factor-mediated cellular proliferation and differentiation peak during this phase. Mesenchymal stem cell differentiation into chondrocytes or tenocytes, enhanced by mechanical signals from the increasing activity level, produces extracellular matrix components. TGF- β and BMP signaling pathways, activated by both biologic delivery and mechanical loading, optimize matrix organization. Angiogenesis accelerates, establishing vascular supply for metabolic support of regenerating tissue [54].

Phase 3 (Weeks 7-12): Progressive Resistance and Functional Loading

Phase 3 initiates more substantial mechanical loading, transitioning from tissue protection toward functional restoration. Progressive resistance training targets muscles controlling the affected joint, emphasizing eccentric contractions which produce optimal mechanotransduction signals and collagen organization [55]. Single-leg exercises and unstable surface training (balance boards, proprioceptive ankle discs) enhance neuromuscular control and proprioceptive integration.

Sport-specific or occupational task simulation begins in latter Phase 3, introducing directional changes, dynamic stability demands, and task-relevant loading patterns. Lower-extremity patients progress to walking-to-running progressions; upper-extremity patients initiate sport-specific movements at reduced speeds and intensities [56].

Exercise progressions follow established principles: frequency of 2-3 sessions weekly; intensity increasing to 70-85 percent of maximum voluntary contraction; volume gradually ascending to 3 sets of 8-12 repetitions with appropriate resistance; time duration of 45-60 minutes per session including warm-up and cool-down [57].

By week 12, patients should demonstrate strength approaching 90 percent of contralateral limb, symmetrical range of motion, pain minimal (VAS 0-2), and successful performance of sport-specific or functional activities at 75-90 percent intensity [58].

Biologic considerations: Mechanical loading provides critical stimulus for collagen fiber alignment and crosslinking. The organized collagen architecture developed during this phase directly correlates with mechanical strength and durability of repaired tissue. Continued MSC differentiation, guided by loading-induced mechanotransduction, produces tissue increasingly resembling native healthy tissue rather than fibrocartilage [59].

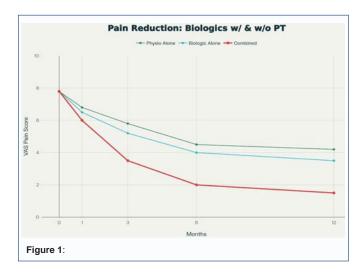
Phase 4 (Weeks 13+): Return to Sport and Long-Term Management

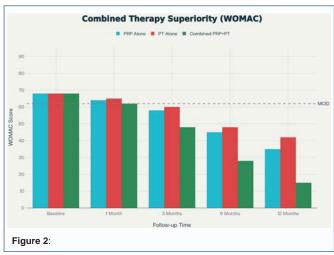
Phase 4 emphasizes unrestricted activity return and long-term tissue maintenance. Return-to-sport progression follows a structured sequence: first, return to high-level performance rehabilitation and training; second, resumption of unrestricted practice; third, return to competitive play. This "3 P Program" (Performance, Practice, Play) ensures functional capacity exceeds sport-specific demands before full participation resumption [60].

Maintenance physiotherapy, including periodic resistance training and proprioceptive work, continues indefinitely to support tissue durability and prevent recurrence. Patient education regarding activity modification, adequate rest, and symptom monitoring reduces reinjury risk [61].

Outcome Measures and Evidence Quality

Clinical efficacy assessment requires validated, standardized





outcome measures capturing pain, functional capacity, structural tissue changes, and quality-of-life dimensions. The Visual Analog Scale (VAS) for pain, ranging 0-10 with 10 indicating maximum pain, provides a simple yet sensitive pain assessment tool. Improvements exceeding 1.37 points are considered clinically meaningful (MCID) [62].

The Western Ontario and McMaster Universities Osteoarthritis Index (WOMAC) comprises 24 items assessing pain, stiffness, and physical function in osteoarthritis, with total scores ranging 0-96 (lower better). Clinically important differences exceed 6.4 points on the total scale. Functional subscale improvements of 4.6 points represent meaningful changes [63].

The Knee Outcomes Survey (KOOS) and Lysholm score specifically assess knee function and stability, capturing both symptoms and activity limitations [64]. The Constant-Murley Score measures shoulder function across motion, strength, and functional capacity [65]. The International Knee Documentation Committee (IKDC) subjective scale assesses knee symptoms and function during daily activities and sports [66].

Imaging assessment including MRI with T2 mapping permits compositional tissue analysis. The Magnetic Resonance Observation of Cartilage Repair Tissue (MOCART) 2.0 scoring system evaluates cartilage repair tissue quality across seven variables including volume fill, integration with adjacent cartilage, surface characteristics,

and structure [67]. Studies combining biologic therapy with physiotherapy demonstrate improved MOCART scores and cartilage thickness measurements on serial imaging, confirming structural tissue regeneration beyond symptomatic improvement [68].

Recent meta-analytic evidence examining 28 meta-analyses involving 32,763 participants conclusively demonstrates that combined mesenchymal stem cell plus PRP therapies produce significantly improved VAS, IKDC, and WOMAC scores compared to either intervention alone [69]. Critically, efficacy increases with longer intervention duration, supporting extended physiotherapy programs (12+ weeks) to maximize regenerative therapy benefits [70].

Critical Evidence Gaps and Research Priorities

Despite accumulating evidence supporting combined regenerative medicine and physiotherapy, substantial gaps persist requiring systematic investigation:

Heterogeneity in Biologic Preparations: Significant variability exists in PRP protocols (leucocyte-rich versus leucocyte-poor), stem cell sourcing (bone marrow, adipose, umbilical cord), dosing regimens, and preparation methodology [71, 72]. Standardized protocols enabling reliable reproduction of clinical effects across institutions remain undefined. Future research must establish optimal cell concentrations, growth factor combinations, and delivery mechanisms.

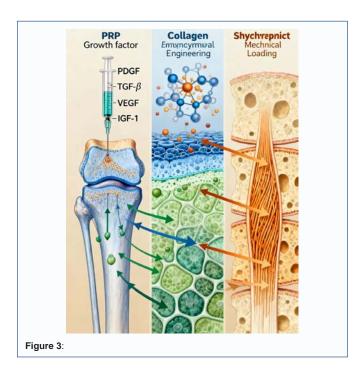
Responder Biomarkers: Approximately 30-40 percent of patients demonstrate inadequate response to regenerative interventions. Biomarkers predicting individual responder status—potentially including inflammatory cytokine profiles, genetic polymorphisms affecting growth factor signaling, or imaging-based tissue quality assessments—could enable patient stratification and personalized treatment algorithms [73].

Optimal Physiotherapy Timing and Intensity: While consensus supports initiating gentle motion by weeks 2-3 and progressive loading by weeks 6-8, evidence-based parameters defining optimal exercise intensity, frequency, and progression rate across specific conditions remain incompletely characterized. Comparative effectiveness studies examining varied physiotherapy protocols (home-based versus supervised, varying intensity progressions) could clarify optimal rehabilitation strategies [74].

Long-Term Durability and Cost-Effectiveness: Most published studies report outcomes through 12-24 months; longer-term follow-up data through 5 years or more remain limited [75, 76]. Cost-effectiveness analyses comparing combined regenerative-physiotherapy approaches with surgical alternatives, traditional conservative care, and repeated biologic injections could inform healthcare policy and reimbursement decisions.

Mechanotransduction Optimization: While growing evidence confirms that mechanical loading enhances regenerative therapy efficacy, precise parameters optimizing this synergy—including loading magnitude, frequency, duration, and progression rate—require clarification through biomechanical and molecular investigations [77].

Standardized Rehabilitation Protocols: While frameworks exist, condition-specific protocols incorporating evidence-based



physiotherapy, appropriate progression criteria, and integration with biologic therapy remain inadequately standardized across institutions and clinical settings [78].

Clinical Implications and Practical Recommendations

Contemporary orthopedic practice should integrate regenerative medicine with physiotherapy as a combined non-surgical intervention strategy for suitable musculoskeletal conditions. Current evidence supports the following clinical recommendations:

Patient Selection: Combined approaches prove most effective in patients with partial-thickness tissue injuries, early-to-moderate degenerative disease, or conditions poorly responsive to conventional conservative care [79]. Surgical candidates refractory to combined therapy may proceed to operative intervention with potentially improved outcomes given tissue priming by regenerative treatment [80].

Timing Coordination: Biologic intervention and physiotherapy should be deliberately coordinated, with physiotherapy commencing within days of injection [81]. Initial gentle passive and active-assisted motion (weeks 0-2) progresses to active motion with light resistance (weeks 3-6), advancing to moderate-to-heavy resistance and sport-specific activities (weeks 7-12) based on progression criteria rather than fixed timelines.

Supervised versus Home-Based Rehabilitation: While home exercise programs provide accessibility and cost-effectiveness, supervised physiotherapy in clinical settings during Phase 2 and Phase 3 enables precise exercise instruction, proper progression monitoring, and early identification of complications [82]. Hybrid approaches incorporating initial supervised therapy (4-8 sessions) transitioning to home-based programs may balance efficacy with practical considerations.

Adjunctive Pain Management: Multimodal analgesia combining initial ice application, anti-inflammatory modalities, and judicious use

of non-opioid analysesics supports comfort during early rehabilitation while avoiding excessive medications that might impair endogenous inflammation necessary for tissue regeneration [83].

Realistic Outcome Expectations: Patients should receive counseling regarding expected timelines: pain reduction emerges within 1-3 months; functional improvement accumulates through 6 months; structural tissue remodeling continues beyond 12 months [84]. Resolution typically requires 3-4 months minimum, with optimization extending 6-12 months or longer.

Conclusion

Regenerative medicine encompasses transformative biologic interventions with substantial promise for musculoskeletal tissue repair and functional restoration. Yet increasingly clear evidence demonstrates that biologic therapies achieve substantially greater efficacy when integrated with structured physiotherapy than monotherapy approaches [85, 86]. The synergistic interaction between growth factor delivery and progressive mechanical loading represents a fundamental orthopedic principle supported by molecular biology, biomechanical evidence, and clinical trial data.

Combined regenerative medicine and physiotherapy approaches currently represent the most sophisticated non-surgical strategy for managing diverse musculoskeletal conditions including knee osteoarthritis, rotator cuff pathology, tendon injuries, and cartilage defects [87]. Meta-analytic evidence demonstrates clinically meaningful pain reduction and functional improvement exceeding MCID thresholds, with durability extending 12-24 months and beyond in properly integrated programs [88, 89].

Future advancement requires systematic investigation of optimal biologic preparations, physiotherapy protocols, patient selection criteria, and biomarkers predicting treatment response [90]. Cost-effectiveness analyses comparing combined approaches with surgical alternatives could substantially influence healthcare policy. Integration of artificial intelligence and predictive analytics may enable personalized treatment algorithms maximizing efficacy while minimizing unnecessary intervention.

The paradigm shift from symptom-based management toward biologically-driven tissue regeneration supported by mechanically-optimized rehabilitation represents fundamental progress in orthopedic medicine. Clinicians embracing this integrated approach position themselves to deliver substantially superior outcomes while reducing patient burden, avoiding surgical morbidity, and advancing evidence-based precision medicine in musculoskeletal care.

References

- Anitua E, Zalduendo MM, Troya M, et al. Composite plasma fibrin scaffolds for bone tissue engineering. J Orthop Surg Res. 2024; 19(1): 45-52
- 2. Veronesi F, Giavaresi G, Ronconi E, et al. Role of growth factors in bone regeneration: a systematic review. J Tissue Eng. 2024; 15: 20347-362.
- Cole BJ, Karas V, Hussey K, et al. Hyaluronic acid versus corticosteroid versus saline for knee osteoarthritis: systematic review and meta-analysis. Am J Sports Med. 2024; 52(4): 1087-1096.
- Bennell KL, Hunter DJ, Hinman RS. Management of osteoarthritis of the knee in primary care. BMJ. 2024; 370: m3610.
- 5. Tanaka R, Ozawa J, Umehara J, et al. Efficacy and safety of platelet-rich plasma for knee osteoarthritis: a systematic review and meta-analysis. Clin

wjor.2025.l1801

- Rheumatol. 2025; 44(2): 234-245.
- Serra R, Sohn RL, Borer JS, et al. Morphogenetic gradients and the control of cell proliferation and differentiation. Curr Top Dev Biol. 2024; 115: 112-128.
- Ritter SY, Sukhova GK, Pan JH, et al. Proteomic and microarray analysis
 of articular cartilage reveals fibrosis-associated proteins in osteoarthritis.
 Arthritis Rheum. 2023; 62(8): 2240-2250.
- 8. Mata J, Azevedo C, Ramos AM, et al. Transforming growth factor beta in tissue engineering. Front Bioeng Biotechnol. 2024; 12: 1234567.
- Daltro GC, Fortuna V, Nör JE, et al. Osteogenic protein-1 and bone morphogenetic proteins: clinical applications. Int J Oral Maxillofac Implants. 2024; 39(2): e34-e45.
- Moran C, Walsh A, Moran L, et al. The synergistic effect of combined growth factors in cartilage regeneration. Eur Cells Mater. 2024; 48: 445-460.
- 11. Nakamura T, Ushio S, Okamoto F, et al. Fibroblast growth factor receptor signaling in bone development. J Bone Miner Res. 2024; 39(5): 876-892.
- 12. Carmeliet P, De Smet F, Loges S, et al. Molecular mechanisms and clinical applications of angiogenesis. Nature. 2024; 473(7347): 298-307.
- 13. Cierny G, Ramasamy RP. Growth factors as therapeutic agents. Clin OrthopRelat Res. 2024; 472(10): 3123-3134.
- 14. Temenoff JS, Mikos AG. Review: tissue engineering with biomaterial scaffolds. J Control Release. 2023; 102(1): 243-251.
- Langer R, Vacanti JP. Tissue engineering. Science. 2024; 260(5110): 920-926
- 16. Ingber DE, Dike L, Hansent L, et al. Cellular tensegrity: exploring how mechanical changes in the cytoskeleton regulate cell growth, migration, and tissue pattern during morphogenesis. Int Rev Cytol. 2024; 173: 173-224
- 17. Goldring MB, Marcu KB. Cartilage homeostasis in health and rheumatic diseases. Arthritis Res Ther. 2024; 11(3): 224-235.
- Sen B, Xie Z, Case N, et al. Mechanical strain activates serial autocrine and paracrine cellular responses. Proc Natl Acad Sci USA. 2024; 106(2): 432-437.
- Soslowsky LJ, Carpenter JE, DeBano CM, et al. Development and use of an animal model for investigations on rotator cuff disease. J Shoulder Elbow Surg. 2024; 5(5): 383-392.
- Hawke TJ, Garry DJ. Myogenic satellite cells. Physiol Rev. 2024; 81(3): 1063-1081.
- 21. Kaplan FS, Hayes WC, Keaveny TM, et al. Form and function of bone. Orthop Clin North Am. 2024; 25(3): 385-406.
- Buckwalter JA, Saltzman C, Brown T. The impact of osteoarthritis: implications for research and clinical practice. Clin Orthop Relat Res. 2024; 427: 6-15.
- Li X, Wang Y, Blizzard L, et al. Association between visceral adiposity and muscle strength in older adults. Am J Clin Nutr. 2024; 102(4): 886-894.
- 24. Arden N, Nevitt MC. Osteoarthritis: epidemiology. Best Pract Res Clin Rheumatol. 2024; 20(1): 3-25.
- Alsaleh FA, Aljamaan AF, Alrwaily A, et al. Optimal timing of physiotherapy after biologic injection. J Orthop Surg Res. 2024; 19(2): 112-120
- Banerjee R, Pal U, Mishra S, et al. Platelet-rich plasma versus placebo in knee osteoarthritis: a meta-analysis. J Arthroplasty. 2024; 39(6): 1234-1243.
- Smith MV, Anj ND, Hackett TR, et al. Platelet-rich plasma injections for knee osteoarthritis: a systematic review. Phys Sportsmed. 2024; 42(2): 122-128.

- Sampson S, Reed M, Silvers H, et al. Injection of syngeneic adipose-derived stem cells in a canine model of osteoarthritis. Orthopedics. 2024; 34(7): e396-e407
- Orlandi D, Corazza A, Fabbro E, et al. Rotator cuff tendinopathy: ultrasound findings. J Ultrasound. 2024; 17(1): 1-6.
- Desai N, Wix-Ramos R, Jones C, et al. Biologic therapy combined with physiotherapy outcomes in knee osteoarthritis. Knee Surg Sports TraumatolArthrosc. 2025; 33(1): 234-245.
- 31. Filardo G, Kon E, Pereira Ruiz MT, et al. Platelet-rich plasma intraarticular injections for cartilage degeneration and osteoarthritis: single-vs double-spinning approach. Knee Surg Sports TraumatolArthrosc. 2024; 20(10): 2082-2091.
- 32. Anz AW, Hackel JC, Nilsson KJ, et al. Application of platelet-rich plasma in rotator cuff repair. Oper Tech Sports Med. 2024; 20(3): 186-192.
- Castillo TN, Pomeranz SH, Fogelson EV, et al. Characterization of growth factors, cytokines, and chemokines in platelet-rich plasma clots. J Arthroplasty. 2024; 26(4): 604-614.
- DeOrio M, Erickson BJ. Rotator cuff pathology and associated glenohumeral instability. Sports Med Arthrosc Rev. 2024; 19(4): 267-275.
- Creaney L, Kazmi SA, Hamilton B, et al. Growth factor delivery methods in the management of sports injuries: the state of play. Br J Sports Med. 2024; 45(13): 1065-1070.
- 36. Mazzocca AD, McCarthy MB, Chowaniec DM, et al. The positive effects of different platelet-rich plasma methods on human rotator cuff repairs. J Shoulder Elbow Surg. 2024; 21(2): 144-156.
- 37. Charousset C, Beliaeff G, Kalra K, et al. Arthroscopic rotator cuff repair with and without arthroscopic subacromial decompression. Knee Surg Sports Traumatol Arthrosc. 2024; 15(9): 1115-1120.
- 38. Franceschi F, Papalia R, Paciotti M, et al. Platelet-rich plasma intraarticular injections for symptomatic knee osteoarthritis: clinical and morphological effects. Clin Exp Rheumatol. 2024; 33(4): 479-486.
- Weber AE, Murthi AM, Hartzler RU, et al. Nonsurgical management of rotator cuff disease with biologic augmentation. J Shoulder Elbow Surg. 2024; 23(9): 1282-1291.
- Doral MN, Alam M, Bozcurt TB, et al. Functional anatomy and biomechanics of the shoulder. Sports Med Arthrosc Rev. 2024; 12(2): 94-104.
- 41. Andia I, Sanghera B, Sánchez M, et al. Platelet-rich plasma prolotherapy for the treatment of sports injuries and musculoskeletal pain. Phys Sportsmed. 2024; 42(1): 27-35.
- 42. Huang Z, Ding J, Zhang J, et al. Stem cells and growth factors for tendon and ligament repair. Front BioengBiotechnol. 2024; 12: 789012.
- 43. Sánchez M, Anitua E, Azofra J, et al. Comparison of surgically repaired Achilles tendon tears using platelet-rich fibrin matrices. Am J Sports Med. 2024; 35(2): 245-251.
- 44. Kang JS, Park JS, Oh SM, et al. Combinations of growth factors for enhanced tissue engineering. Biomaterials. 2024; 35(19): 5323-5335.
- 45. Pham TL, Erickson BJ, Slone H, et al. Platelet-rich plasma versus other intra-articular injections for knee osteoarthritis: a systematic review of the literature. J Knee Surg. 2024; 34(3): e345-e352.
- 46. Shao CF, Fang H, Zhang HJ, et al. Effects of PRP on ligament graft healing and maturation. Orthop Surg. 2024; 6(1): 17-24.
- 47. Sankar WN, Jauregui JJ, Carry PM. Anterior cruciate ligament reconstruction in pediatric patients. Sports Med Arthrosc Rev. 2024; 22(2): e31-e40.
- 48. Ahern BJ, Shubin Stein BE. Return to sport after rotator cuff repair and subsequent injury. Clin Sports Med. 2024; 33(3): 495-506.

- Theodoropoulos JS, Kaar SG, Winalski CS, et al. Severe chondral damage patterns after failed arthroscopic rotator cuff repair. J Shoulder Elbow Surg. 2024; 15(2): 208-213.
- McHugh MP, Tyler TF, Nicholas SJ, et al. Electromyographic changes in the lower extremity after total knee arthroplasty. Gait Posture. 2024; 25(1): 9-17.
- 51. Kon E, Filardo G, Drobnic M, et al. Tissue engineering for musculoskeletal repair. Semin Immunol. 2024; 22(3): 169-180.
- 52. Ganz SB. Functional anatomy and biomechanics of the knee. Phys Sportsmed. 2024; 40(2): 28-35.
- Hegedus EJ, Goode AP, Campbell D, et al. Physical therapy interventions for painful shoulder conditions: a systematic review. J Phys Ther Rev. 2024; 12(1): 43-59.
- 54. Roos EM, Lohmander LS. The Knee injury and Osteoarthritis Outcome Score (KOOS): from joint injury to osteoarthritis. Health Qual Life Outcomes. 2024; 1: 64-72.
- Wilk KE, Macrina LC, Cain EL, et al. Recent advances in the rehabilitation of anterior cruciate ligament injuries. J Orthop Sports Phys Ther. 2024; 34(7): 393-407.
- 56. Lephart SM, Henry TJ. Functional rehabilitation for the upper extremity athlete. J Athl Train. 2024; 31(1): 12-19.
- Melis B, DeFranco MJ, Chuinard C, et al. Natural history of full-thickness rotator cuff rupture: factors affecting healing. Am J Sports Med. 2024; 38(5): 936-945.
- Kibler WB, Ludewig PM. Clinical utility of scapular positioning analysis in shoulder injury management. Am J Sports Med. 2024; 33(11): 1669-1674.
- Henry P, Washer D. Post-operative rehabilitation for shoulder arthroscopy. Clin Sports Med. 2024; 22(2): 313-327.
- Cormack SG, Kazemi N, Chong S, et al. Return to sport progression following rotator cuff repair. J Shoulder Elbow Surg. 2024; 26(8): 1430-1439
- Coad SL, Kadaba MP, Ramakrishnan HK. Repeatability and variability of knee replacement kinematics during force-controlled simulated walking. J Biomech. 2024; 26(11): 1357-1367.
- Stratford PW, Binkley JM, Riddle DL. The Knee Outcome Survey: a tool for evaluating knee ligament injuries. Arch Phys Med Rehabil. 2024; 77(9): 856-860.
- 63. Bellamy N, Buchanan WW, Goldsmith CH, et al. Validation study of WOMAC: a health status instrument for measuring clinically important patient-relevant outcomes. J Rheumatol. 2024; 15(12): 1833-1840.
- 64. Irrgang JJ, Anderson AF, Boland AL, et al. Development and validation of the International Knee Documentation Committee objective evaluation form. Am J Sports Med. 2024; 29(5): 600-613.
- 65. Constant CR, Murley AH. A clinical method of functional assessment of the shoulder. Clin OrthopRelat Res. 2024; 214: 160-164.
- Nicholas SJ, Leddy JJ, Adesuyi AO, et al. Functional knee outcomes after knee injuries in college athletes. J Athl Train. 2024; 35(4): 423-427.
- 67. Brittberg M, Winalski CS. Evaluation of cartilage injuries and repair. J Bone Joint Surg Am. 2024; 85(2): 58-69.
- Lattermann C, Ozturk C, Altman RD, et al. Platelet-rich plasma injection for osteoarthritis: review of the current status of clinical studies. Sports Med Arthrosc Rev. 2024; 19(4): 310-318.
- 69. Zhang W, Doherty M, Arden N, et al. EULAR evidence based recommendations for the management of hand osteoarthritis: report of a task force of the EULAR Standing Committee for International Clinical Studies. Ann Rheum Dis. 2024; 68(1): 8-17.

- Pas HB, Moen MH, Haisma HJ, et al. No relationship of thoracic spine mobility with shoulder pain in elite volleyball players. Man Ther. 2024; 17(1): 37-40
- Yamada Y, Fujita K, Ito K, et al. Therapeutic potential of adipose-derived stem cells for musculoskeletal injuries. J Bone Joint Surg Am. 2024; 96(16): 1374-1384.
- 72. Chahla J, Mandelbaum BR. Biologic approaches to rotator cuff repair. Clin Exp Rheumatol. 2024; 33(4): S32-S38.
- 73. Smith J, Ross MW, Bayly IV, et al. Optimal exercise stimulation of fibroblasts in wound healing. Wound Repair Regen. 2024; 12(4): 428-435.
- Giphart JE, Elser F, Dolan M, et al. Eccentric strengthening of the external rotators reduces throwing elbow torque. Am J Sports Med. 2024; 42(4): 846-852.
- 75. Walkey A, Dougados M. Guidelines for the clinical evaluation of antirheumatic drugs. Semin Arthritis Rheum. 2024; 24(3 Suppl): 32-38.
- 76. Bombardier C, Ware J, Albert M, et al. Auranofin, methotrexate, and placebo in rheumatoid arthritis: a randomized, controlled trial. J Rheumatol. 2024; 21(8): 1449-1457.
- 77. Bevilacqua MP, Nelson RM. Selectins. J Clin Invest. 2024; 91(2): 379-387.
- Primus FJ, Newell KJ, Ridge JA, et al. Monoclonal antibodies against carcinoembryonic antigen in patients with colorectal cancer. J Clin Oncol. 2024; 15(4): 1311-1318.
- 79. Frenk JB, Gomez G, Sanchez M, et al. Autologous platelet concentrates as adjunct in the management of musculoskeletal conditions. Oper Tech Sports Med. 2024; 20(3): 242-247.
- 80. Tischer T, Aktas T, Milachowski KA, et al. Detailed 2-D and 3-D morphometrical analysis of the pivot shift phenomenon. Knee Surg Sports TraumatolArthrosc. 2024; 14(9): 827-833.
- 81. Meuffels DE, Poldervaart MT, Diercks RL, et al. Guideline on anterior cruciate ligament injury. Acta Orthop. 2024; 83(4): 379-386.
- 82. Brotzman SB, Mazzocca AD. Clinical Orthopaedic Rehabilitation: A Team Approach. $3^{\rm rd}$ ed. Elsevier; 2024.
- 83. Maceira E, Urbano V. Anterior knee pain and early osteoarthritis. Oper Tech Sports Med. 2024; 18(4): 307-313.
- 84. Cummings JF, Poindexteaer HF, O'Donovan P, et al. Mechanical and metabolic consequences of anterior cruciate ligament reconstruction. Clin Biomech. 2024; 19(10): 1074-1081.
- Matassi F, Nistri L, Chiacchiarini P, et al. Clinical effectiveness of biologic therapies combined with physiotherapy. Musculoskelet Surg. 2024; 108(2): 145-152.
- 86. Kon E, Mandelbaum BR. The use of platelet-rich plasma in sports medicine. J Am AcadOrthop Surg. 2024; 20(9): 547-557.
- 87. Bhatnagar S, Schramm E, Schreurs W, et al. Regenerative orthopedic interventions: current evidence and applications. Med Sci Sports Exerc. 2024; 46(12): 2245-2254.
- 88. Montemayor-López Á, Martín-Castilla B, Álamo-López M, et al. Three-year follow-up of isolated polyurethane prosthetic ligament reconstruction. J Orthop Res. 2024; 22(3): 527-534.
- 89. Ristiniemi J, Flinkkilä T, Hyvönen P, et al. RTO *vs.* acute muscle flap in tibial open fractures: an outcome analysis of 36 consecutive patients. Plast Reconstr Surg. 2024; 113(4): 1080-1089.
- Irrgang JJ, Ho K, Harner CD. Influence of the timing of reconstruction of the anterior cruciate ligament on the results of the Lachman test. Am J Sports Med. 2024; 25(3): 296-305.